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Original article

# Investigation of the bone metabolism in dogs with leishmaniasis

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## Abstract

The present study was aimed at determining bone metabolism related changes in dogs with visceral leishmaniasis such as vitamin D, calcium, phosphate and ALP levels as compared to healthy ones. A total of 40 serums of male dogs aged between 4 and 6 years that were sent to Adnan Menderes University, Faculty of Medicine, Parasitology Laboratory from Kuşadası and Bodrum districts between the years 2012 and 2013, suspected with leishmaniasis, by the veterinary physicians in accordance with the cold chain rules and 20 of which were evaluated as leishmania seropositive and 20 as leishmania seronegative by the IFA test, were included in this study. There was no any statistically significant difference between the serum 25-OH-D<sub>3</sub> levels of Leishmania positive and negative dogs ( $p>0.05$ ). There was a statistically significant difference between the serum ALP ( $p<0.05$ ), and P levels were observed to be higher in the seropositive dogs than in the seronegative dogs, while Ca level was low ( $p<0.001$ ).

**Key words:** leishmaniasis, vitamin D, calcium, phosphate, alkaline phosphatase

## Introduction

Leishmaniasis is an important zoonotic infectious disease which can be observed in many countries of the world. The disease is prevalently observed in the Mediterranean Region, Africa, Asia and Central Southern America (Papadopoulou et al. 2005). While it is especially observed in the coastal regions in Turkey, cases from nearly all of the regions were reported (Özbel et al. 2002).

Dogs are the most important reservoir host for *Leishmania infantum*, which is the agent of the visceral disease (Oliva et al. 2010). Depending on the immune response of the animals which contracted the disease, the course of the disease can be asymptomatic or symptomatic (Desjeux et al. 2004). The pri-

mary symptoms observed on the skin can include ash hyperkeratosis, skin dehydration, scleroderma, loss of elasticity, depigmentation, scurfy dermatitis (Gönül et al. 2002). Furthermore, anemia, neutropenia, thrombocytopenia, uremia, hyperproteinaemia, hypergammaglobulinemia, hyperalbuminemia, an increase in ALT, GGT, LDH and ALP levels, in urea and creatine level can be listed as mild or severe proteinuria laboratory findings. The clinical findings may show variability depending on the course, phase and severity of the disease in Canine visceral leishmaniasis (CVL) (Freitas et al. 2012, Taher et al. 2015).

Calcium and phosphorus are important in terms of the development of bone structure and preservation of mineral density. These are also the most frequently found elements in the organism and they have

important physiologic functions in many biological mechanisms, except for the skeletal system (Kalaycıoğlu et al. 2000, Pineda 2003). There are hormonal control mechanisms for the preservation of blood calcium and phosphorus levels. These mechanisms consist of 3 hormones, which are parathyroid hormone (PTH), calcitonin and vitamin D (1,25 dihydroxycholecalciferol). These hormones have also an effect on the bone and mineral metabolism (Oleszek and Marston 2000, Fidan and Dundar 2007).

Vitamin D is a group sterol with hormone-like functions. The most important effect of vitamin D is its effect on calcium homeostasis. It is responsible for the bone metabolism, passing of the calcium ions, which are necessary for neural functions in the bone structure, through the cell membrane,  $\text{Ca}^{+2}$  absorption from the intestines and mobilization from the bones (Kalaycıoğlu et al. 2006). The main activation stage in the vitamin D metabolism takes place in the kidneys and is catalyzed by 25-hydroxycalciferol-1 $\alpha$ -hydroxylase. 1,25 dihydroxycholecalciferol (1,25 dihydroxyvitamin D) formed as a result of this hydroxylation is the most active form of vitamin D and it is responsible for all the effects of vitamin D (Gow et al. 2011, Dittmer and Thompson 2011).

Although the metabolic function of alkaline phosphatase is not completely understood, it is considered to play an important role in the intestinal lipid transport and calcification in the bones (Onat et al. 2002, Murray et al. 2004). It is found on the outer surfaces of absorptive cells and it helps the transport process. It is considered to be an enzyme that regulates the active transport *in* the cell membrane (Uzunoğlu 1998).

Alkaline phosphatase is found in the liver, bone tissue, kidneys, intestines, placenta, pancreas, thyroid gland, tooth enamel, leucocyte cell membrane, endoplasmic reticulum, mitochondria, nuclear membrane and cell membrane (Uzunoğlu 1998). Although its metabolic function is not completely understood, it is considered to have an important role in the intestinal lipid transport and calcification in the bones and regulates the active transport in the cell membrane (Onat et al. 2002, Murray et al. 2004). The alkaline phosphatase activity is one of the most frequently used biochemical indications in bone diseases and it is partially caused by osteoblasts. Thus, in osteoporotic patients, serum alkaline phosphatase activity reaches higher values when compared to the normal course (Çetiner et al. 2001, Ecer et al. 2005).

Changes in the alkaline phosphatase level observed in the laboratory findings in dogs with leishmaniasis suggest that leishmaniasis may cause bone formation (Çetiner et al. 2001, Ecer et al. 2005). In this study, in order to determine the effect of canine

leishmaniasis on the bone metabolism, serum calcium, phosphate, alkaline phosphates and 25-OH-D<sub>3</sub> values, among the biochemical indicators of the bone, were investigated.

## Materials and Methods

### Samples

A total of 40 serums of male dogs aged between 4 and 6 years that were sent to Adnan Menderes University, Faculty of Medicine, Parasitology Laboratory from Kuşadası and Bodrum districts in Turkey between the years 2012 and 2013, suspected with leishmaniasis, by the veterinary physicians in accordance with the cold chain rules, and 20 of which were evaluated as leishmania seropositive and 20 as leishmania seronegative by the Indirect Immunofluorescence Antibody Test (IFAT), were included in this study.

### Determination of anti-leishmania antibodies by the indirect antibody fluorescence method

Leishmania antibodies were studied by the IFAT method in the serum samples. In the study, rabbit anti-dog IgG fluorescein isothiocyanate conjugated (Sigma, F-7884) was used at 1/100 dilution.

The samples were analyzed under the fluorescence microscope (Olympus CH-40) with an excitation filter at 490 nm wavelength, with 40 mm lenses by using a 20-mm ocular. In the study, 1/64 and above titers were evaluated as positive.

### Biochemical Analyses

The indication of the calcium amount in the blood serum with the glyoxal-bis method was measured by the colorimetric method. This method is based on the measurement made by using the calcium chelator feature of glyoxal-bis in the presence of other +2 valent cations (Bellinger and Campbell 1965).

The serum inorganic phosphate level was determined as reductive in the presence of tin chloride, by the colorimetric method based on the formation of ammonium molybdiphosphate the reaction of phosphate ions with ammonium molybdate in an acidic medium (Davies et al. 1973). The analyses of serum alkaline phosphatase (Spinreact®, Catalogue no: BEIS44-I) and 25-OH-D<sub>3</sub> (Immunodiagnostic Systems Ltd, Catalogue no: UK51081) were made by using a kit.

Table 1. Visceral leishmaniasis seropositivity in the dogs according to the IFAT results.

Titers	Seropositive dog	
	number	%
1/64	1	5
1/128	3	15
1/256	4	20
1/512	12	60
Total	20	100

Table 2. Vitamin D, ALP, Ca and P results investigated in the dog serums.

	Leishmaniasis negative (n:20)	Leishmaniasis positive (n:20)	p
25-OH-D <sub>3</sub> nmol/L	70.01 ± 14.12	51.89 ± 6.59	0.252
ALP U/L	104.5 ± 8.7	137.1 ± 11	0.026*
Ca mg/100ml	8 ± 0.5	3.38 ± 0.48	0.000*
P mg/100ml	2.19 ± 0.17	3.01 ± 0.11	0.000*

## Statistical Analyses

For the statistical analysis of the data obtained, SPSS (for Windows Release 15.0 Standard Version-Copyright© Spss Inc. 1989-2001) package program was used. The significance analyses were performed using the Mann-Whitney U Test.

## Results

The titer rates of 20 Leishmania-positive male dogs that were included in the study are shown in Table 1.

Serum 25-OH-D<sub>3</sub>, ALP, Ca and P results of Leishmania positive and Leishmania negative male dogs are given in Table 2.

There was not any statistically significant difference between the serum 25-OH-D<sub>3</sub> levels of Leishmania positive and negative dogs ( $p > 0.05$ ). There was statistically significant difference between the serum ALP ( $p < 0.05$ ) and P levels were observed to be higher in the seropositive dogs than in the seronegative dogs while Ca level was low ( $p < 0.001$ ).

## Discussion

Leishmaniasis is among the most neglected diseases in the world and it is reported by the World Health Organization (WHO) that the disease is endemic in a total of 98 countries, 350 million people are at risk, and there are estimated 2 million new cases in a year. The mortality and morbidity of leishmaniasis

show a tendency to increase every year at an alarming rate (WHO 2010). Two epidemiologic forms, zoonotic and anthroponotic, are observed in leishmaniasis. The frequently observed form in the Mediterranean countries is the zoonotic form. The form observed in East Africa, Bangladesh and India is the anthroponotic form (Molano et al. 2003).

Many different methods are used in the diagnosis of canine leishmaniasis. These methods can be listed as cytohistological methods, serological tests, parasitological methods, molecular methods and determination of the cellular immune response. The sensitivity and selectivity of the IFAT method are reported to be close to 100%, and it is acknowledged as the reference serological method by the World Organization for Animal Health (WOAH) (Mettler et al. 2005).

The researchers reported that the IFA technique was a quite sensitive and specific test for the determination of leishmaniasis, and upon evaluating the clinical findings and laboratory results together, the titrations between 1/40 and 1/1024 were diagnostic in seropositive patients and the titers above 1/40 were sufficient for the diagnosis of canine leishmaniasis (Freitas et al. 2012). In our study, titers of 1/64 and above are accepted as positive.

In the investigation of the bone metabolism, vitamin D, alkaline phosphatase, Ca and P are generally evaluated together. It has been shown in the investigations that vitamin D is closely related to calcium and phosphorus homeostasis, and plays a significant role in the bone model formation (Dittmer and Thompson 2011). In the study of Driel et al. (2006) conducted in order to investigate the autocrine/paracrine function of 1 $\alpha$ , 25-OH-D<sub>3</sub>, it was determined that 1 $\alpha$ -hy-

droxylase is expressed in the human osteoblasts, and also there are megalin and cubilin protein receptors bound to vitamin D. It was observed that osteoblasts produced  $1\alpha, 25\text{-}(\text{OH})_2\text{D}_3$ , and ALP and osteocalcin expressions increased mildly with the addition of  $25\text{-OH-D}_3$  as a substrate to the osteoblast cell cultures while it did not have any effect on Ca and parathyroid hormone regulation. In the studies conducted on humans,  $25\text{-OH-D}_3$  showed a relation with osteoporosis,  $25\text{-OH-D}_3$  levels were observed to be low in patients with osteoporosis, while the parathormone levels were high (Sahota et al. 1999, Sahota et al. 2004, Garneo et al. 2007). The vitamin D metabolism in dogs was mostly investigated in the pathological cases of kidney failure (Galler et al. 2011). In the literature no study investigating the vitamin D metabolites in leishmania-infected dogs was encountered.

In this study conducted with the purpose of investigating the effect of leishmaniasis on the bone metabolism in dogs, serum  $25\text{-OH-D}_3$  levels in infected dogs were found to be lower than those in healthy dogs. However, this difference is statistically insignificant.

In the study conducted by Rallis et al. (2005), from 26 dogs infected with *Leishmania infantum*, anemia in 21 dogs, an increase in the total amount of protein in 23 dogs, low albuminemia in 6 dogs, an increase in the ALP activity in 15 dogs and hyperbilirubinemia in 2 dogs were determined. Similarly, in the studies conducted on dogs with leishmaniasis, the levels of ALP, ALT, SGOT, LDH, total protein, blood urea nitrogen, creatinine, creatine kinase, and cholesterol were observed to be higher than the normal values (Tyrphonas et al. 1977, Koutinas et al. 2001). In the present study, the serum ALP levels in the dogs naturally infected with leishmania were determined to be high similarly to the findings of other researchers.

Paşa et al. (2003) found that serum zinc and iron levels in the dogs with visceral leishmaniasis were significantly lower than those in healthy dogs, while the copper level was high. Any significant differences in the calcium, phosphorus, and magnesium values were not observed. A positive correlation between hyperphosphatemia and the severity kidney failure was observed in 155 dogs with leishmaniasis at various stages of chronic kidney failure (Cortadellas et al. 2009). Upon evaluating the studies in general, it is observed that hypocalcemia and hyperphosphatemia occur in the cases in which kidney failure develops along with leishmaniasis. In the study conducted, the P and Ca results suggest that kidney failure may be found in the infected dogs.

As a result, the facts that clinical findings of leishmaniasis are not always clear and the symptoms vary depending on the immune system in patients make

the clarification of the pathophysiology of the disease difficult. Upon evaluating the results in this study, which the bone metabolism in dogs with visceral leishmaniasis investigated, in general, an increase in the ALP level suggests that the findings of hypocalcemia and hyperphosphatemia may have adverse effects on the bone metabolism.

## References

- Cortadellas O, Fernandez Del Palacio MJ, Talavera J, Bayon A (2009) Serum phosphorus homeostasis in dogs with spontaneous chronic kidney disease at different stages severity. *J Vet Intern Med* 24: 73-79.
- Cetiner S, Ozturk M, Yucetas S (2001) Determination of serum alkaline phosphatase, calcium and phosphate levels after the clinical use of solvent dehydrated allogenic bone implantation in cystic cavities. *Hacettepe Diş Hekimliği Fakültesi Dergisi* 25: 26-30.
- Coskun S, Batmaz H, Aydyn L, Yılmaz F (1997) Seroprevalence of *L. infantum* infection of dogs in the western part of Turkey. *Türkiye Parazitoloj Derg* 21: 287-291.
- Desjeux P (2004) Leishmaniasis: Current Situation and New Perspectives. *Comp Immunol, Microbiol Infect Dis* 27: 305-18.
- Dittmer KE, Thompson KG (2011) Vitamin D Metabolism and Rickets in Domestic Animals. *Vet Pathol* 48: 389-407.
- Driel M, Koedam M, Buurman CJ, Hewison M, Chiba H, Uitterlinden AG, Pols HA, Van Leeuwen JPM (2006) Evidence for auto/paracrine actions of vitamin D in bone: 1-hydroxylase expression and activity in human bone cells. *FASEB* 20.
- Ecer S, Dikici B, Hasbolat K (2005) Bone Mineral Metabolism in Chronic Liver Diseases. *Dicle Tıp Dergisi* 32: 57-62.
- Fidan AF, Dundar Y (2007) The Hypocholesterolemic and Antioxidant Effects of *Yucca schidigera* and Its Saponins and Phenolic Matters (A Review) *Lalahan Hay Araşt Enst Derg* 47: 31-39.
- Freitas JC, Nunes-Pinheiro DC, Neto BE, Santos GJ, Abreu CR, Braga RR, Campos RM, Oliveira LF (2012) Clinical and laboratory alterations in dogs naturally infected by *Leishmania chagasi*. *Rev Soc Bras Med Trop* 45: 24-29.
- Galler A, Tran JL, Krammer Lukas U, Holler U, Thalhammer JG, Zentek J, Willman M (2011) Blood vitamin levels in dogs with chronic kidney disease. *Vet J* 192: 226-31.
- Garnero P, Munoz F, Sornay-Rendu E, Delmas PD (2007) Associations of vitamin D status with bone mineral density, bone turnover, bone loss and fracture risk in healthy postmenopausal women. The OFELY study. *Bone* 40: 716-722.
- Gow AG, Else R, Evans H, Berry JL, Herrtage ME, Mellanby RJ (2011) Hypovitaminosis in dogs within inflammatory bowel disease and hypoalbuminaemia. *J Small Anim Pract* 52: 411-418.
- Gonul R, Arun SS, Dodurka T, Handemir E (2002) *Leishmania infantum* in a dog. *Turk J Vet Anim Sci* 26: 689-694.

- Kalaycıoğlu L, Serpek B, Nizamlioğlu M, Başpınar N, Tiftik AM (2000) Biochemistry 2<sup>nd</sup> edition. Ankara: Nobel Yaynevi p: 35-36.
- Kalaycıoğlu L, Serpek B, Nizamlioğlu M, Başpınar N, Tiftik AM (2006) Biochemisrty. Nobel Yayyn Dağıtım, Ankara p: 55.
- Kouzinis K, Saridomichelakis MN, Mylonakis ME, Leontidis L, Polyzopoulou Z, Billinis C, Argyriadis D, Diakou N, Papadopoulos O (2001) A randomised, blinded, placebo-controlled clinical trial with allopurinol in canine leishmaniosis 98: 247-261.
- Lainson R, Shaw JJ (1987) Evolution, classification and geographical distribution. In Peters W, Killick-Kendrick R, editors. The Leishmaniasis in Biology and Medicine, vol. Orlando: Academic Press p: 1-120.
- Mettler M, Grimm F, Capelli G, Camp H, Deplazes P (2005) Evaluation of enzyme-linked immunosorbent assays, an immunofluorescent-antibody test, and two rapid tests immunochromatographic dipstick and gel tests) for serological diagnosis of symptomatic and asymptomatic Leishmania infections in dogs. J Clin Microbiol 43: 5515-5519.
- Molano I, Alonso MG, Mirón C, Redondo E, Requena JM, Soto M, Nieto CG, Alonso C (2003) A *Leishmania infantum* multi-component antigenic protein mixed with live BCG confers protection to dogs experimentally infected with *L. infantum*. Vet Immunol Immunopathol 92: 1-13.
- Murray RK, Granner DK, Mayes PA, Rodwell VW (2004) Harper's Illustrated Biochemistry Translators: Mentis G, Ersoz B. Nobel Typ Kitabevleri Istanbul.
- Oleszek W, Marston A (2000) Saponins in Food, Feedstuffs and Medicinal. Kluwer Academic Publishers p: 260.
- Oliva G, Roura X, Crotti A, Maroli M, Castagnaro M, Gradoni L, Lubas G, Paltrinieri S, Zatelli A, Zini E (2010) Guidelines for treatment of leishmaniasis in dogs. J Am Vet Med Assoc 236: 1192-1198.
- Onat T, Sönmez E, Emek K (2002) Human Biochemistry Palme Yayincılık p: 213-220.
- Ozbel Y, Oskam L, Ozensoy S, Turgay N, Alkan MZ, Jaffe CL, Ozcel MA (2000) Epidemiology of canine leishmaniasis in western Turkey: Comparison of serological, molecular, biological and parasitological procedures. Acta Trop 74: 1-6.
- Ozbel Y, Turgay N, Alkan MA, Babaoglu A, Ozensoy S, Babalyoglu N (2002) A zoonotic visceral leishmaniasis focus: Karabuk in Western Black Sea Region. Turkiye Parazitoloji Dergisi 26: 362-366.
- Papadopoulou C, Kostoula A, Dimitriou D, Panagyou A, Bobojianni C, Antoniadis G (2005) Human and canine leishmaniasis in asymptomatic and symptomatic population in Northwestern Greece. J Infect 50: 53-60.
- Pasa S, Kargin F, Bildik A, Seyrek K, Ozbel Y, Ozensoy S (2003) Serum and Hair Levels of Zinc and other elements in dogs with visceral leishmaniasis. Biological Trace Element Research 94: 141-147.
- Pineda MH (2003) Mc Donald's Veterinary Endocrinology and Reproduction. Iowa State Press p:102.
- Rallis T, Day MJ, Saridomichelakis MN, Adamama-Moraitou KK, Papazoglou L, Fytianou A, Koutinas AF (2005) Chronic Hepatitis Associated with Canine Leishmaniasis (*Leishmania infantum*): a clinicopathological study of 26 Cases. J Comp Path 132: 145-152.
- Sahota O, Masud T, San P, Hosking J (1999) Vitamin D insufficiency increases bone turnover markers and enhances bone loss at the hip in patients with established vertebral osteoporosis. Clin Endocrinol 51: 217-221.
- Sahota O, Munday MK, San P, Godber IM, Lawson N, Hosking DJ (2004) The relationship between vitamin D and parathyroid hormone calcium homeostasis, bone turnover, and bone mineral density in postmenopausal women with established osteoporosis. Bone 35: 312-319.
- Taher JH, Abdullah NA, Muhammed S, Faris E (2015) Evaluation of Some Enzymes Levels in Iraqi Children Infected with Visceral Leishmaniasis. Scholar Research Library, Der Pharma Chemica 7: 1-5.
- Tryphonas L, Zavidzka Z, Bernard MA, Janzen EA (1977) Visceral leishmaniasis in a dog: Clinical, hematological and pathological observations. Can J Comp Med 41: 1-12.
- Uzunoeu N (1998) Physicochemical properties of alkaline phosphatase enzyme. T Klinikleri Typ Bilimleri 18: 69-75.
- WHO (2010) Control of leishmaniasis, World Health Organ Tech Res Ser, 2010/01/01ed: 1-86. [http://apps.who.int/iris/bitstream/10665/44412/1/WHO\\_TRS\\_949\\_en.pdf](http://apps.who.int/iris/bitstream/10665/44412/1/WHO_TRS_949_en.pdf)