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Original article

CpG islands: Features and distribution in the genomes of porcine parvovirus

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Abstract

Porcine parvovirus disease is a reproductive disorder caused by the porcine parvovirus (PPV) in sows and is characterised by miscarriage, stillbirth and mummification in pregnant sows. Porcine parvovirus disease poses a significant threat to pork herds and seriously hinders healthy and sustainable development of the pig farming industry. Currently, there is no effective treatment for porcine parvovirus disease except for prevention and control measures. Based on genotype differences, PPV can be classified into at least eight subtypes, PPV1-PPV8. Epigenetic mechanisms, particularly cytosine methylation of cytosine-phosphate-guanine (CpG) dinucleotides, are proven to have a significant impact on the life cycle of various viruses. Therefore, we selected the PPV genome as the research object and analysed the number, distribution and length of CpG islands in the genome of strains PPV1-PPV8. PPV1-6 had AT rich genomes (GC content $\leq 50\%$), whereas PPV7 had a GC content >50%. PPV1, PPV4, PPV5 and PPV6 contained fewer CpG islands (1-5), PPV7 contained moderate CpG islands (6-11) and PPV2 and PPV3 contained more CpG islands (12-16). This study provides a foundation for exploring novel antiviral treatment strategies from an epigenetic perspective.

Keywords: CpG islands, DNA methylation, porcine parvovirus

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Introduction

Parvoviruses infect a wide range of hosts, including vertebrates, invertebrates and humans (Streck et al. 2024, Vargas-Bermudez et al. 2023). According to the different host types, the family Parvoviridae can be classified into two subfamilies: Parvovirinae and Densovirinae, which infect vertebrates and invertebrates, respectively (Cotmore et al. 2019, Penzes et al. 2020). The subfamily of parvoviruses consists of eight virus genera, namely, the Aleutian mink disease virus (Amdoparvovirus), avian parvovirus (Aveparvovirus), Bocaparvovirus (Bocaparvovirus), replicativirus (Copiparvovirus), dependent virus (dependent virus), erythrovirus (erythroparvovirus), protoparvovirus (Protoparvovirus) and tetraparvovirus (Vargas-Bermudez et al. 2023). In 1965, Mayr and Mahnel first discovered the porcine parvovirus (PPV), which is one of the main pathogens that causes reproductive disorders in sows (Mayr et al. 1968). In 1967, Cartwright et al. (Cartwright et al. 1969). isolated porcine parvovirus from disease samples when conducting etiological research on infertility, abortion and stillbirth in pigs, thus proving the pathogenic effect of PPV for the first time. Following viral infection in the pre-pregnancy period, the virus invades the embryo or foetus through the placenta, causing abortion, embryo death, foetal malformation, foetal mummification and infertility in sows; however, the sows themselves have no apparent clinical symptoms (Meszaros et al. 2017). Porcine parvovirus may play a role in diarrhoea, dermatitis and respiratory diseases, in addition to coinfection with porcine reproductive and respiratory syndrome virus, porcine circovirus type 2 and other common viruses (Chen et al. 2023, Faustini et al. 2024). In recent years, rising cases of PPV infections have caused marked economic losses to the pig industry.

PPV virus particles are hexagonal or circular, without a capsule, with a diameter of 20-23 nm and an icosahedral equiaxed stereosymmetry (Streck and Truyen 2020, Vargas-Bermudez et al. 2023). The PPV genome is a single-stranded linear DNA, with a size of 4-6 kb and palindromic sequences at both ends (Streck and Truyen 2020, Vargas-Bermudez et al. 2023). Three structural proteins are encoded by the PPV genome: VP1, VP2 and VP3 (Ranz et al. 1989, Vargas-Bermudez et al. 2023). VP1 and VP2 overlap in many amino acid sequences (Streck and Truyen. 2020, Vargas-Bermudez et al. 2023). VP1 is a structural protein necessary for PPV replication and viral particle packaging, and functions to stabilise viral single-stranded DNA (Streck and Truyen 2020, Vargas-Bermudez et al. 2023). Additionally, the VP1 N-terminus contains a proline-rich region that plays an important role in extracellular to intracellular viral transfer, and VP1 has an N-terminus-rich alkaline residue that helps to bind with the single- -stranded DNA, thereby initiating the replication and packaging of viral DNA (Streck and Truyen 2020, Vargas-Bermudez et al. 2023). VP2 is the main component of capsid proteins and exhibits a haemagglutinating activity. The peptide encoded by VP2 can self- -assemble into virus-like particles, therefore functioning as an effective antigen transporter that induces strong cellular immunity (Streck and Truyen 2020, Vargas-Bermudez et al. 2023). VP3 is a product of the post-translational cleavage processing of VP2 and appears only after capsid assembly and viral genome packaging (Simpson et al. 2002). The PPV genome encodes three non-structural proteins: NS1, NS2 and NS3 (Streck and Truyen 2020, Vargas-Bermudez et al. 2023). The NS1 protein is the main non-structural protein involved in PPV infection, and its genes are highly conserved (Streck and Truyen 2020). However, the specific function of this encoded protein remains unclear. To date, there are eight known genotypes of PPV that can infect pigs: PPV1-8 (Vargas-Bermudez et al. 2023). PPV1 can cause reproductive disorders in sows that clinically manifest as miscarriages, mummified foetuses and stillbirths, causing substantial economic losses to the pig farming industry (Streck and Truyen 2020). PPV2-8 is a new type of porcine parvovirus that has been reported both domestically and internationally in recent years (Jager et al. 2021). Currently, research on these new types of porcine parvoviruses is focused on epidemiological studies (Li et al. 2021) thus, their pathogenicity and mechanisms remain unknown.

DNA methylation is a form of DNA chemical modification that can alter genetic expression without altering the DNA sequence (Moore et al. 2013, Mattei et al. 2022). Cytosine methylation, which occurs only in eukaryotes, refers to the covalent modification of the methyl group on the 5th carbon atom of cytosine in cytosine-phosphate-guanine (CpG) dinucleotides (under the catalytic action of DNA methyltransferases (DNMTs) to form 5-methylcytosine (5mC)) (Moore et al. 2013, Mattei et al. 2022). DNA methylation often occurs in CpG sequence density-enriched regions; namely, CpG islands (CGIs). CGIs exist in three regions: transcription initiation sites, gene bodies and transposable elements (Moore et al. 2013, Mattei et al. 2022).

DNA methylation plays a crucial role in embryonic and germ cell development (Isagawa et al. 2011). Research has found that early embryonic and germ cell development in mammals involves overall demethylation and re-establishment of methylation maps (Isagawa et al. 2011). The target tissues for PPV infec-

B. Schematic representation of CpG isolation from NADL-2. C. Schematic representation of the CpG isolate in the MZ706996 strain. The red circle represents the CpG island in the genome.

tion are the reproductive organs, causing subsequent reproductive disorders (Vargas-Bermudez et al. 2023). PPV infection in the early stages of pregnancy (30-50 days) causes foetal abortion and sows exhibit recurrent oestrus and infertility, whereas infection during mid-pregnancy (50-60 days) can cause stillbirth, mummified foetuses and malformed foetuses (Joo et al. 1976). Infection at 70 days of pregnancy can lead to miscarriage and stillbirth (Joo et al. 1976). In the late stages of pregnancy (after 70 days), most infected pigs give birth normally; however, the resulting piglets are small and have a high risk of mortality (Joo et al. 1976). Piglets that survive can still carry the virus. Therefore, different stages of infection may result in varying clinical symptoms. In this preliminary study, we explored the biological characteristics of viruses from the perspective of DNA methylation, laying the foundation for correlation research between epigenetics and viral pathogenicity.

Materials and Methods

Sequencing information

PPV1-PPV8 sequences were collected from the GenBank of National Biotechnology Information Center, using the search terms 'Porcine Parvovirus' and 'complete genome'.

CpG island analysis

We conducted a CpG island analysis of the genome sequences of PPV1-PPV8. Two online software packages were used: Meth Primer (http://www.urogene.org/ cgi-bin/methprimer/methprimer.cgi) and CpG Plot (www.ebi.ac.uk/Tools/seqstats/embass _cpg) to calculate the CpG islands of the PPV genome sequences for each genotype. CpG islands were calculated by detecting the GC content and observing the expected CpG dinucleotide ratio of a sequence window. The standard settings for CpG islands were as follows: sequence window length should not be <100 bases, GC content should not be <50%, and observation/expected value should not be <60%.

Results

CGI distribution in PPV1

The search results indicated that PPV1 (with a known full-length genome sequence) includes three strains: MZ577027.1, MZ577026.1 and MZ706996.1. Using the MZ706996.1 strain as an example, the total genome length was 4494 bp. The predicted results indicated that the sequence contained a CpG island located between positions 1898 and 1999 with a length of 102 bp (Fig. 1C). In addition, strains with known sequences (including the NADL-8, NADL-2, and Kresse strains) were analysed. The results indicated that the Kresse strain contained a CpG island located at positions 4384-4490 bp, with a length of 106 bp (Fig.1A). The NADL-8 and NADL-2 strains did not contain CGI (Fig.1B).

CGI distribution in PPV2

The analysisshowed that there are a total of 19 strains of PPV2 with known full-length genome sequences (Fig. 2A and Table 1) with a GC content of 44-45% containing 12-16 CGI. Using the MW051675 strain as an example, the total genome length was 5324 bp. The prediction results indicated that the sequence con-

Fig. 2. GC and CpG content of porcine parvovirus 2 (PPV2) genomes. A. The value of GC percentage and CpG island in PPV2 genomes. Left vertical bars represent the value of the CpG island in the genomes. Right vertical bars represent value of GC content. Light blue box indicates the representative strains analysed. B. Schematic representation of the CpG island in the MW051675 strain virus. Red circles symbolize the CpG island in the genomes.

tained 12 CGI located at positions 50-435, 1096-1198, 1273-1444, 1544-1848, 1910-2208, 2235-2499, 2563-2781, 2999-3657, 3859-3981, 4058-4249, 4341-4479, and 4757-5266 bp, with lengths ranging from 103 to 659 bp (Fig. 2B).

CGI distribution in PPV3

The results indicated that a total of eight PPV3 had known full-length genome sequence (Fig. 3A and Table 2), with a GC content of 50% containing 12-14 CGI. Taking the KU167029 strain as an example, its genome length was 5081 bp. The prediction results showed that the sequence contained 12 CGI located at positions 49-215, 763-1164, 1358-1481, 1562-1895, 2142-2276, 2526-2641, 2790-3024, 3121-3224, 3276-3378, 3398-4450, and 4760-4935 bp, with lengths ranging from 103 to 523 bp (Fig. 3B).

Fig. 3. GC and CpG content of porcine parvovirus 3 (PPV3) genomes. A. The value of GC percentage and CpG islands in PPV3 genomes. Left vertical bars represent the value of CpG islands in the genomes. Right vertical bars represent value of GC content. Light blue box indicates the representative strains analysed. B. Schematic representation of the CpG island in the KU167029 strain virus. Red circles symbolize the CpG island in the genomes.

CGI distribution in PPV4

A total of 14 PPV4 exhibited known full-length genome sequences (Fig. 4A and Table 3), of which 12 sequences contain 1-3 CGI s with an average GC content of 41%. Taking the MH921911 strain as an example, the total genome length was 5479 bp. The prediction results indicated that there is a CpG island located at positions 593-694 bp, with a length of 102 bp (Fig. 4B).

CGI distribution in PPV5

The retrieval results showed that PPV5 with a known full-length genome sequence included 16 strains (Fig. 5A and Table 4), with a GC content of 39% and 1-3 CGI s. The JX896318 strain was used as an example, the total genome length was 5516 bp, which contained two CGI s, located at positions 49-159 and 3165-3292 bp, respectively, with lengths of 111 and 128 bp, respectively (Fig. 5B).

CGI distribution in PPV6

We identified a total of 38 known full-length genome sequences of PPV6 strains (Fig. 6A and Table 5), with an average GC content of 45-46% and containing 3-5 CGI s. For example, MH447537has a total genome length of 5976 bp. and four CGI s located at positions 826-941, 1721-1821, 2046-2168 and 5036-5161 bp, with lengths ranging from 101 to 126 bp (Fig. 6B).

CGI distribution in PPV7

The search results indicated that PPV7, which has a known full-length genome sequence, included 44 strains (Fig. 7A and Table 6) with a GC content of 53-55% containing 6-11 CGI s. Using the MZ577044

Fig. 4. GC and CpG content of porcine parvovirus 4 (PPV4) genomes. A. The value of GC percentage and CpG islands in PPV4 genomes. Left vertical bars represent the value of CpG islands in the genomes. Right vertical bars represent value of GC content. Light blue box indicates the representative strains analysed. B. Schematic representation of the CpG islands in the MH921911 strain virus. Red circles symbolize CpG islands in the genomes.

strain as an example, the total genome length was 3999 bp. The prediction results indicated that the sequence contained seven CGI s located at positions 48-178, 389-897, 1158-1899, 1932-2463, 2537-2767, 2819-3393 and 3665-3832 bp, respectively, with lengths ranging from 168 to 742 bp (Fig. 7B).

Discussion

PPV has only one serotype that can be classified into four categories based on pathogenicity and tissue preference: The first type is represented by the NADL-8 strain, which is highly virulent (Meszaros et al. 2017). Oral administration can cross the placental barrier, leading to foetal infection and the formation of viremia. The second type (represented by NADL-2) is an attenuated vaccine strain that cannot cross the placental barrier after oral administration, is non-pathogenic to pregnant sows and foetuses and does not cause viraemia (Jozwik et al. 2009). Therefore, the second type can be used as an attenuated vaccine strain. When the NADL-2 strain is inoculated into the uterus, the virus has the ability to replicate and infect, which can lead to foetal death. The third type (represented by the Kresse strain) is a highly virulent strain of dermatitis that can kill piglets with insufficient immunity (Meszaros et al. 2017). The fourth type is enteritis type strains, which mainly cause intestinal lesions (Csagola et al. 2016).

Fig. 5. GC and CpG content of porcine parvovirus 5 (PPV5) genomes. A. The value of GC percentage and CpG islands in PPV5 genomes. Left vertical bars represent the value of CpG islands in the genomes. Right vertical bars represent value of GC content. Light blue box indicates the representative strains analysed. B. Schematic representation of the CpG islands in the JX896318 strain virus. Red circles symbolize CpG islands in the genomes.

Reports have shown that PPV DNA was found to be methylated independently from its origin (Toth et al. 2013). Like the encapsidated negative strand, the positive strand has been shown to be hypomethylated, thus indicating that PPV DNA maintains hypomethylation throughout the entire lifecycle of the virus, including replication and packaging (Toth et al. 2013). This study showed that the PPV1 strain Kresse contains a CpG isolate, whereas NADL-8 and NADL-2 do not. NADL-8 may be the reason for the short analysis sequence, which was only 3000 bp. The PPV NADL-2 strain contained 60 CpGs. According to previous reports, single nucleotide polymorphisms (SNPs) in the PPV genome were analysed based on available sequences from DNA databases, and the CpG site in the PPV genome was found to be more variable than the GC or C and G sites (Toth et al. 2013). This finding supports the high mutation rate of CpG sites in the PPV genome, which may explain the low number of CGIs.

Previous studies have shown that methylation strat-

egies for combating viruses include methylation of the viral genome and demethylation of the host genome (Gao et al. 2021, Verdikt et al 2022). This study analyses the potential methylation sites of various types of PPV genomes and explores the methylation characteristics of PPV from an epigenetic perspective. The function of methylation is to shut down gene activity, and

the function of demethylation is to activate gene expression (Einkauf et al. 2022, Rehman et al. 2023). Once the DNA and RNA of a virus are methylated, their replication, transcription, and translation activities will inevitably decrease (Einkauf et al. 2022, Rehman et al. 2023). The implementation of DNA methylation requires the involvement of DNA methyltransferase

Accession no.	Length (bp)	Location/Length (bps)	Location/Length (bps)	Location/Length (bps)	Location/Length (bps)	Location/Length (bps)
KY094494	6199	933-1046	1826-1926	2151-2273	5142-5266	
MZ577040	6400	84-251	1096-1209	1989-2089	2314-2436	5305-5429
MZ577039	6199	933-1046	1826-1926	2151-2273	5142-5266	
MH921913	5490	818-933	1713-1813	2038-2160	3254-3374	5029-5153
MH921909	5967	821-939	1719-1819	2044-2166	5035-5159	
MH921907	5974	824-939	1719-1819	2044-2166	5034-5159	
MH921906	5965		1719-1819	2044-2166	5035-5159	
MH921903	5965		1712-1813	2038-2160	5029-5153	5681-5784
MH921901	5994	836-951	1731-1831	2056-2178	5046-5171	
MH921900	5959	818-933	1713-1813	2038-2177	5029-5153	
MH447541	5940	818-933	1713-1813	2038-2160	3254-3374	5029-5153
MH447540	5959	818-933	1713-1813	2038-2177	5029-5153	
MH447539	5996	838-953	1733-1833	2058-2180	5048-5173	
MH447538	5967		1721-1821	2046-2168	5037-5161	
MH447537	5976	826-941	1721-1821	2046-2168	5036-5161	
MH447536	5969	823-941	1721-1821	2046-2168	5037-5161	
MH447535	5979	839-952	1732-1832	2057-2179	5048-5172	5700-5803
MG760726	6123	856-971	1751-1851	2076-2198	5067-5191	
NC 023860	6148	856-971	1751-1851	2076-2198	5066-5191	
KR709268	6148	858-971	1751-1851	2076-2198	3292-3412	5067-5191
KR709267	6117	856-971	1751-1851	2076-2198	5067-5191	
KR709266	6144	858-971	1751-1851	2076-2198	3292-3412	5067-5191
KR709265	6144	858-971	1751-1851	2076-2198	3292-3412	5067-5191
KR709264	6143	858-971	1751-1851	2076-2198	5067-5191	
KR709263	6128	856-971	1751-1851	2076-2198	5066-5191	
KR709262	6147	858-971	1751-1851	2076-2198	5067-5191	
KF999685	6148	856-971	1751-1851	2076-2198	5066-5191	
KF999684	6163	858-971	2076-2187	4532-4643	5072-5191	
KF999683	6148	858-971	2076-2197	4532-4643	5073-5191	
KF999682	6136	858-972	2076-2187	4532-4643	5072-5191	
KF999681	6136	858-972	2076-2187	4532-4643	5072-5191	
MW853957	6571	943-1056	1836-1936	2161-2283	5152-5276	
MW853956	6287	65-216	2279-2396	4737-4847	5275-5394	
MW853955	6309	86-237	2300-2417	4758-4868	5296-5415	
MW853954	6328	51-222	1059-1174	1954-2054	2279-2401	5270-5394
MW051672	6382	977-1090	2195-2317	3411-3531	5164-5310	
MK825573	6180	2145-2251	4601-4712	4602-4713	5141-5260	
MH558679	6144	858-971	2076-2198	3292-3412	5068-5191	

Table 5. Locations and lengths of CpG islands within genome of PPV6.

(DMT), a methyl donor, and methylation of specific bases (Rehman et al. 2023). On the 35th day of pregnancy, PPV infection leads to embryonic death (Joo et al. 1976). At approximately 70 days of pregnancy, the clinical symptoms of foetal infection are obscure (Joo et al. 1976). Embryonic development undergoes cell reprogramming, allowing an individual cell to transition to a pluripotent embryo or a pluripotent stem cell (Isagawa et al. 2011, Han et al. 2024). During this process, the genomic DNA sequence remains unchanged (Isagawa et al. 2011, Han et al. 2024). Although this process is short, the changes in cell fate and gene expression regulation are very drastic. Early embryos can observe a wide range of highly methylated states Table 6. Locations and lengths of CpG islands within genome of PPV7.

in both transcriptional and non-transcriptional regions (Han et al. 2024). And there may not be excess, readily available methyl groups to methylate the invading viral DNA. This may be the molecular biological basis for the rapid development of early embryos into death, which is not closely related to the type of virus. Can future corresponding viral therapies utilize this mechanism, especially viral DNA methylation, to suppress viral pathogenicity? At least this is a direction worth further research.

This study extended the analysis of the number and distribution of CpG islands in the PPV2-PPV7 genome (no data available for PPV8). The results indicated differences in the number and distribution of CpG islands. Research has shown that methylation of CpG islands can impair transcription factor binding, recruit inhibitory methyl-binding proteins and stably silence gene expression (Modutlwa et al. 2009). Although the PPV1 genome maintains low methylation independent of the tissue of origin throughout the virus lifecycle, further research is needed to investigate the methylation status of the PPV2-PPV7 genome and determine the extent to which DNA methylation of CpG islands regulates gene expression. Other studies have demonstrated that CpG islands, particularly those associated with gene promoters, are rarely methylated (Zhao et al. 2009). In addition, previous research has found that. The NADL-2 strain of PPV contains 60 CpGs but does not have a CpG island (Toth et al. 2013). Toll-like receptor 9 (TLR9) is a member of the Toll-like receptor family, which can be activated by non-methylated cytidine monophosphate guanosine DNA (CpG DNA) derived from pathogens or artificially synthesised oligonucleotides containing non-methylated CpG (CpG ODN) and directly or indirectly initiates innate immune responses through downstream signalling, thereby resisting pathogen invasion (Kim et al. 2013). Recent studies have shown that parvoviruses do not induce immune responses activated by TLR9 (Mattei et al. 2013). Therefore, further clarification is needed to determine whether methylation occurs at the CpG site outside the CpG island in the PPV genome and its role.

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